

Thermodynamic stability of folded proteins against mutations

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By balancing the average energy gap with its typical change due to mutations for protein-like heteropolymers with M residues, we show that native states are unstable to mutations on a scale $M^* \sim (\lambda/\sigma_\mu)^{1/\zeta_s}$, where λ is the dispersion in the interaction free energies and σ_μ their typical change. Theoretical bounds and numerical estimates (based on complete enumeration on four lattices) of the instability exponent ζ_s are given. Our analysis suggests that a limiting size of single-domain proteins should exist, and leads to the prediction that small proteins are insensitive to random mutations.

In order for a protein to perform a biological activity it has to fold to a structure with a well-defined topology. It is widely believed that this native state has the lowest free energy [1]. It is less well appreciated that the folded state of a single-domain protein is only marginally stable compared to the ensemble of unfolded structures [2]. Thus the polypeptide chain can undergo large amplitude fluctuations after reaching the native conformation. Another characteristic of single-domain proteins is that they are small: the number of amino acid residues in single-domain proteins seldom exceeds 200. Larger proteins usually self-organize into multi-domain structures [2].

The marginal stability of single-domain proteins suggests that polypeptide chains may be sensitive to mutations that alter the primary sequence of amino acids. On the other hand, many different sequences often lead to similar native states, i.e., sequence homology gives rise to a class of folded states which are topologically similar [2]. Thus, from the perspective of homologous proteins we expect that point mutations may have little effect on the stability of the folded states of proteins. These observations raise the question: to what extent are native states of proteins stable when subjected to random mutations?

Experimentally, the stability of folded states against random uncorrelated mutations can only be examined by synthesizing a library of mutated sequences starting from a known structure [3]. In this article we investigate the stability of folded states using a combination of theoretical arguments and exact enumeration for various lattice models of proteins.

Since our arguments are general, we can only quantify the effect of mutations in statistical terms. In particular, we have analyzed how the probability that a mutation causes a change in the native state of a protein depends on the number of residues. We assume that the conformations in the native and first excited state are not structurally related, so that it is reasonable to focus on these two states to estimate the effect of random mutations. For the simplified models discussed below we have verified this assumption by explicit computation of the overlap between the two states for a number of sequences. For real proteins the gap would correspond to

the free energy difference between the native basin of attraction (NBA) and the closest competing basin of attraction (CBA); the structures in the NBA and CBA are expected to be dissimilar as well.

The main result of our analysis is that the energy gap Δ_{av} , averaged over a large number of sequences, and the typical change σ_Δ in the energy gap due to mutations scale with the number of residues M as

$$\Delta_{av} \sim \lambda M^{-\theta}, \quad \sigma_\Delta \sim \sigma_\mu M^\alpha, \quad (1)$$

with $\alpha, \theta > 0$, where λ is the dispersion of the interaction free energies along the chain, and σ_μ is the typical change in interaction free energy for each residue due to mutations. Since Δ_{av} decreases with M while σ_Δ increases, it follows that there is a crossover length M^* , scaling with the instability exponent $\zeta_s = \alpha + \theta$ as

$$M^* \sim (\lambda/\sigma_\mu)^{1/\zeta_s}. \quad (2)$$

The native state is robust with respect to mutations if $M \ll M^*$, but there is a considerable chance that the native state is destabilized by mutations if $M \gtrsim M^*$.

An estimate for α defined in Eq. (1) goes as follows. Let n_Δ denote the number of contacts that are different between the ground state and the first excited state. The independent mutations in the interaction free energies, of typical size σ_μ for each residue, combine to give a change in the gap energy of magnitude $\sigma_\Delta \sim \sigma_\mu \sqrt{n_\Delta}$. Assuming that the n_Δ different contacts are located on a fractal surface, we obtain the inequality $\alpha \geq (d-1)/2d$. The maximum possible number of contacts, which can only occur for $d = \infty$, is $n_\Delta^{\max} = M^2$. Thus $(d-1)/2d \leq \alpha < 1$.

To motivate why Δ_{av} decreases algebraically with M consider an ensemble of n random energies, drawn from a Gaussian distribution $f(E) \propto \exp(-E^2/2)$. The distribution function for the energy gap Δ between the two smallest values among the n samples is given by $f(\Delta) \propto n \int_{-\infty}^{\infty} dE f(E) [g(E + \Delta)]^{n-1}$, where $g(E) = \int_E^{\infty} dE' f(E')$ equals the probability that the energy is larger than E . It turns out (see also below) that $f(\Delta)$ is very well described by an exponential distribution function, $f(\Delta) = (1/\Delta_{av}) \exp(-\Delta/\Delta_{av})$. Consequently, we have $1/\Delta_{av} = -(d/d\Delta) \log f(\Delta)|_{\Delta=0} =$

$-n \int_{-\infty}^{\infty} dE E f(E) [g(E)]^{n-1}$. By numerically evaluating this integral for various values of n we find that for large n the average gap decreases as $\Delta_{\text{av}} \sim (\log n)^{-\theta}$ with $\theta \simeq 0.2$ in the range $10 \leq n \leq 10^6$. Since different conformations may have considerable overlap, the effective number n of independent samples is smaller than the total number of conformations, which grows exponentially with M . But it is reasonable to assume that n still depends exponentially on the number of residues, so that $n \sim c^M$ gives rise to $\Delta_{\text{av}} \sim M^{-\theta}$. This power law behavior is independent of the constant c .

To numerically verify the scaling behavior of Eq. (1), which leads to the length scale M^* (cf. Eq. (2)), we have used simplified lattice models for proteins [4] that allow for complete enumeration of all conformations for chains of moderate length. In these models, a protein is represented by a self-avoiding walk on a lattice. Natural proteins are made from twenty amino acids, roughly half of which are hydrophobic. Contacts between hydrophobic residues which form the core of the protein are favored. To mimic the diversity of natural proteins we use a model in which each residue is assigned a random “charge”, λ_j , drawn from a Gaussian distribution with average $\langle \lambda_j \rangle = \lambda_0$ and variance $\sqrt{\langle (\lambda_j - \lambda_0)^2 \rangle} = \lambda$ [5]. Positive λ_j ’s may be identified as hydrophilic, whereas negative values of λ_j correspond to hydrophobic residues. As a typical value for the ratio of the average hydrophobicity and its dispersion we use $\lambda_0/\lambda = -0.1$. This choice is consistent with the fact that in nature approximately 55% of the residues are hydrophobic [6]. We have used this value in most of our calculations, but as will be shown below, the results are rather insensitive to λ_0 in the range $-1 < \lambda_0/\lambda < 0$.

We use four different lattices: the square and the triangular lattice in two dimensions, and the diamond and the cubic lattice in three dimensions. Consider a conformation ν in which the M residues in the chain reside on lattice sites \mathbf{r}_j^ν , with j labeled 1 through M . By definition, a contact occurs between two residues j and k if they are nearest neighbors on the lattice. In terms of a (symmetric) contact matrix C a contact is represented by $C_{jk}^\nu = 1$. Subsequent residues j and $j+1$ are always nearest neighbors on the lattice, and therefore $C_{j,j+1}^\nu \equiv 0$. We also define $C_{jj}^\nu \equiv 0$.

When two residues j and k are in contact, they give a contribution to the total energy E^ν that is by definition equal to the average, $E_{jk} = (\lambda_j + \lambda_k)/2$, of their “charges”. Thus, the total energy of a conformation is given by

$$E^\nu = \sum_{j < k} C_{jk}^\nu \left(\frac{\lambda_j + \lambda_k}{2} \right) = \sum_j n_j^\nu \left(\frac{\lambda_j}{2} \right), \quad (3)$$

where $n_j^\nu = \sum_{k \neq j} C_{jk}^\nu$ equals the number of contacts in which residue j is involved. For end point residues, n_j takes values from 0 to $z-1$, where z is the coordination

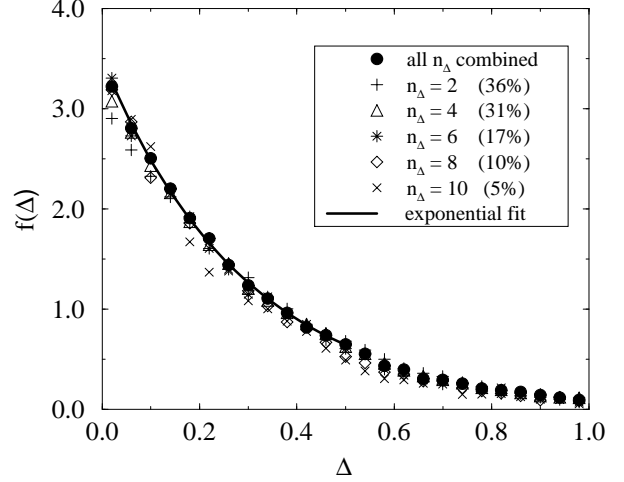


FIG. 1. Distribution of energy gap Δ between native and first excited state for the diamond lattice model with $\lambda_0 = -0.1$ and $M = 18$, obtained using an ensemble of 10^5 sequences. The solid line denotes an exponential fit to the data shown as filled circles. The other symbols correspond to a restriction to specific values of n_Δ . The percentages indicate the fraction of sequences for which each n_Δ occurs.

number of the lattice; for internal residues the maximum number of contacts equals $z-2$.

It is important, both from a theoretical and a computational point of view, that the energy of a conformation ν depends on the coordinates $\{\mathbf{r}_j^\nu\}$ only through the contact matrix $\{C_{jk}^\nu\}$, and in fact an even more reduced description in terms of $\{n_j^\nu\}$ is possible. Therefore, as long as we are interested in thermodynamic rather than kinetic properties, it is sufficient to consider all possible contact sets $\{n_j^\nu\}$. For each value of M we enumerated all possible conformations $\{\mathbf{r}_j^\nu\}$, determining $\{n_j^\nu\}$ for each conformation, and only adding it to the list if it is different from all previously generated contact sets.

The list thus generated was used to find the energies of the native and the first excited state for a large number of random sequences. Many different spatial conformations $\{\mathbf{r}_j^\nu\}$ can give rise to the same set of pair contacts $\{n_j^\nu\}$, which makes our approach very efficient. To quantify this, we first note that the total number of different conformations, C_M , grows exponentially with M , as $C_M \sim z_{\text{eff}}^M$, where z_{eff} is an effective coordination number [7]. The number of contact sets C'_M also grows exponentially with M , but with an effective coordination number $z'_{\text{eff}} < z_{\text{eff}}$, so that $C'_M/C_M \sim (z'_{\text{eff}}/z_{\text{eff}})^M$. On the cubic lattice $z_{\text{eff}} \simeq 4.684$ [7] while $z'_{\text{eff}} \simeq 3.4$, and for $M = 15$ the contact set enumeration is already more efficient by a factor of 400. The description of the energies in terms of the variables $\{n_j^\nu\}$ therefore allows us to calculate the low energy states for a much larger number of sequences than would otherwise have been possible.

Consider the ground state ($\nu = 0$) and first excited

state ($\nu = 1$) for a given sequence. The energy gap, $\Delta = E^1 - E^0$, is defined as the energy difference between the native state and the first excited state. A histogram of Δ , obtained by determining the native state and first excited state for 10^5 randomly drawn sequences $\{\lambda_j\}$ fits very well to the exponential distribution (see Fig. 1).

The mutations are modeled by adding a small perturbation to the “charges”, $\lambda_j \rightarrow \lambda_j + \mu_j$, where μ_j is a Gaussian random variable with zero mean and variance σ_μ . Other ways of simulating the effect of mutations in lattice models have also recently been considered [8]. The effect of a mutation depends on the degree of overlap between $\{n_j^0\}$ and $\{n_j^1\}$. Mutations affecting contacts that occur in both the native state and the first excited state have no influence on the energy gap Δ , since E^0 and E^1 are changed by the same amount. When a contact is not shared between the two states however, mutations will affect the energies of both states differently. To quantify the effect of the mutations in terms of $\{n_j^0\}$ and $\{n_j^1\}$ we consider the effective number of different contacts,

$$n_\Delta = \sum_{j=1}^M (n_j^1 - n_j^0)^2. \quad (4)$$

As a result of the mutations the energy gap is changed by an amount equal to the sum of n_Δ independent Gaussian variables $\mu_j/2$, $\Delta \rightarrow \Delta + \mu_\Delta$, where μ_Δ is a Gaussian variable with zero mean (this is a direct consequence of the fact that $\langle \mu_j \rangle = 0$) and variance $\sigma_\Delta = \sigma_\mu \langle \sqrt{n_\Delta} \rangle / 2$. The values of n_Δ^2 for an ensemble of sequences $\{\lambda_j\}$ are exponentially distributed for large enough M .

Since Δ and σ_Δ are both derived from the same native state and first excited state, it is important to analyze to what extent they are correlated. To this end we have plotted in Fig. 1 the distribution of the energy gap Δ for subsets of sequences for which n_Δ has a particular value. It can be seen that the distribution is essentially independent of n_Δ . Therefore we are justified in considering Δ and n_Δ , or equivalently Δ and σ_Δ , to be independent.

Figure 2 shows how Δ_{av} and σ_Δ depend on the number of residues for the square and the diamond lattice models: σ_Δ/σ_μ is an increasing function of M , while Δ_{av}/λ decreases with M . This behavior holds qualitatively for all four lattices studied. Power law fits to the data for large M , shown as solid lines in Fig. 2, provide estimates for the exponents α and θ defined in Eq. (1) for various values of λ_0/λ . In the range $-0.5 < \lambda_0/\lambda < 0$ there is no significant dependence on λ_0/λ . For $\lambda_0/\lambda = 0.5$, when the chain is largely hydrophilic, α and θ have different values, although their sum (and consequently the instability exponent $\zeta_s = \alpha + \theta$) hardly changes. However, in the limit $\lambda_0/\lambda \rightarrow -\infty$, where only maximally compact states contribute, the scaling behavior breaks down completely, and the dependence on M is quite erratic for the whole range of M values considered.

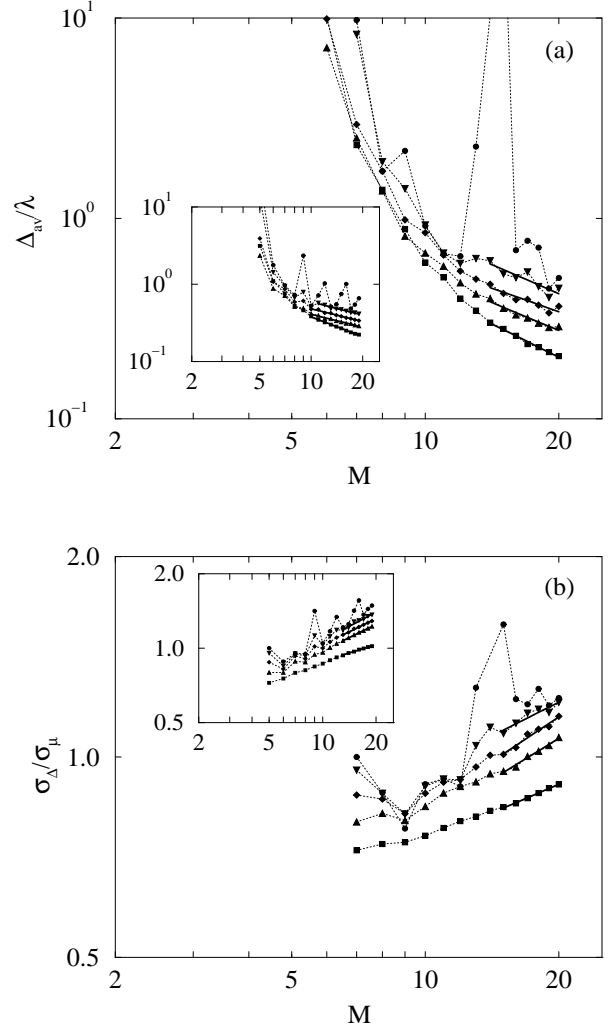


FIG. 2. Dependence of (a) the average energy gap Δ_{av} , and (b) the typical change in the gap due to mutations σ_Δ , on the number of residues M for the diamond lattice model. The insets show the same data for the square lattice model. Different symbols denote various values of λ_0/λ , from top to bottom: $-\infty$, -1.0 , -0.5 , -0.1 , and 0.5 . The data were obtained by averaging over 40,000 random sequences.

On all four lattices we find $\alpha = 0.3 \pm 0.1$, which is consistent with the theoretical bound $(d-1)/2d \leq \alpha < 1$ in both two and three dimensions. We find $\theta = 0.6 \pm 0.2$ in two dimensions (square and triangular lattice) and $\theta = 1.0 \pm 0.2$ in three dimensions (diamond and cubic lattice). Both these values are significantly larger than the theoretical prediction $\theta \simeq 0.2$. It is quite possible that in the range of M values that allow for complete enumeration, θ is enhanced by finite-chain-length effects, and that for larger M the decay of $\Delta_{\text{av}}/\lambda$ is slower and θ reaches its predicted value $\theta \simeq 0.2$. But for M^* as defined in Eq. (2) to exist we only require that $\theta > 0$. The values of the instability exponent $\zeta_s = \alpha + \theta$ for the various lattices are 0.9 ± 0.1 (square), 1.0 ± 0.2 (triangular), 1.2 ± 0.2 (diamond), and 1.3 ± 0.3 (cubic). A more accurate determination of the exponents would require much larger values of M .

In Ref. [9] it was shown that an energy function like the one defined in Eq. (3) — i.e. a sum of one-residue properties rather than a sum of pair interactions — provides a good parametrization of the interaction matrix that was constructed by Miyazawa and Jernigan [10] by statistically analyzing a large number of known protein structures obtained from the Protein Data Bank. To verify this claim explicitly we have performed calculations using the interaction matrix of Ref. [10] and find values for the exponents that are in the same approximate range as the values obtained using the “charge” model [11].

In obtaining the above results for random heteropolymers we have averaged over all sequences without regard to the degeneracy of the ground state. Since the native states of proteins are thought to be unique [1], we have repeated our calculations by including only those sequences that have a non-degenerate native state. Surprisingly, the values of α and θ , and hence of the instability exponent ζ_s , are virtually unchanged.

It might be argued that for our results to be applicable to proteins, which in the course of evolution may have been “optimized” (e.g. with respect to folding kinetics [12]), one must only include “optimized” sequences in the averaging process. Although this may change the value of the instability exponent ζ_s , we expect that α and θ will both remain positive. This expectation is reasonable given the fact that our results are virtually unchanged when we consider only non-degenerate ground states, as mentioned above. Thus we expect our conclusion that the native state is unstable against weak mutations for sufficiently long chains also to hold for (single-domain) proteins.

Since in biological systems mutations occur at a given rate and the dispersion in the interaction free energies of the residues has a limited range of values [2], the fact that single-domain proteins with more than about 200 residues rarely occur may be explained by the sensitivity of the native state to small random perturbations when the number of residues is large. To make this more pre-

cise, consider $p_d = \int_0^\infty d\sigma \int_0^\sigma d\Delta f(\Delta|\Delta_{\text{av}})g(\sigma|\sigma_\Delta)$, i.e., the probability that the folded state is destabilized by the mutations, which only depends on the ratio $x \equiv \sigma_\Delta/\Delta_{\text{av}}$. Here $f(\Delta|\Delta_{\text{av}})$ is an exponential distribution with average Δ_{av} and $g(\sigma|\sigma_\Delta)$ is a Gaussian distribution with zero mean and width σ_Δ . For small x we have $p_d \sim x$ so that using Eq. (2) we obtain $p_d = c(M/M^*)^{\zeta_s}$ for $M \ll M^*$, with c a constant. Now let ε be a small threshold probability separating stable ($M < M_\varepsilon$) from unstable ($M > M_\varepsilon$) proteins, where M_ε is defined as the value of M for which $p_d = \varepsilon$. Eliminating M^* in favor of M_ε we obtain

$$\frac{p_d}{\varepsilon} = \left(\frac{M}{M_\varepsilon} \right)^{\zeta_s}. \quad (5)$$

Given a pair $(\varepsilon, M_\varepsilon)$ we can therefore predict the stability for other chain lengths $M \lesssim M_\varepsilon$ using Eq. (5), provided that we know the value of the instability exponent ζ_s . Note that the ratio of the p_d values for two different values of M is independent of ε , and is thus a suitable quantity to be considered in experiments designed to verify Eq. (5). Since $\zeta_s \simeq 1$, the destabilization probability p_d decreases more or less linearly with M . It follows that on average small proteins are relatively insensitive to point mutations.

In order to utilize our theory for analyzing experiments, it should be kept in mind that the models provide a coarse-grained description of a protein, where each bead corresponds to b amino acids, with $3 \lesssim b \lesssim 4$ [13]. In terms of the number of amino acids M_{aa} and the free energy dispersion per amino acid λ_{aa} , the corresponding model parameters are $M = M_{\text{aa}}/b$ and $\lambda = \lambda_{\text{aa}}\sqrt{b}$. The coarse-graining does not affect Eq. (5).

It is important to estimate the effect of thermal fluctuations on the predictions of our theory. According to Eq. (1), for large enough M we have $\Delta_{\text{av}} = c_\Delta \lambda M^{-\theta}$, with $c_\Delta \simeq 3$ and $\theta \simeq 1$ obtained from the power law fits of Fig. 2. Taking into account the coarse-graining factor b and estimating $\lambda_{\text{aa}} \simeq (15-25)kT$, we find for medium-sized proteins with $M_{\text{aa}} \simeq 100$ that $\Delta_{\text{av}} \simeq (3-5)kT$. We conclude that thermal fluctuations do not significantly affect our conclusions as summarized by Eq. (2). Note that the scaling of Δ_{av} with M suggests that smaller proteins in general have larger melting and denaturation temperatures [14].

There are some striking similarities between our findings and certain results probing the chaotic nature of the spin-glass phase. Using an Imry-Ma style argument [15], Bray and Moore [16] showed that in the ground state of a disordered Ising model the average excitation energy for a cluster of linear size L scales as L^y , while the typical energy associated with random bond perturbations scales as $L^{d_S/2}$, with $d_S/2 > y$. As a consequence, at sufficiently large length scales the ground state is unstable against arbitrarily weak perturbations to the bonds. In

this sense, the ground state of spin-glasses is “chaotic” [16–18], and by analogy the same could be said about the native state of a protein.

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